

# Synthesis and *in vitro* enzymatic and antiviral evaluation of phosphoramidate d4T derivatives as chain terminators

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The anti-HIV activity of nucleoside analogues is highly related to their substrate specificity for cellular and viral kinase and, as triphosphate, for HIV-RT. A series of phosphoramidate d4T derivatives have been synthesized and evaluated as substrates for HIV-1 RT, and also tested for their *in vitro* anti-HIV activity. Compounds **2** and **4** are able to inhibit HIV-1 replication to the same extent as d4T and d4TMP in MT-4 cells as well as in CEM/0 cells and CEM/TK<sup>-</sup> cells. The data suggests that these phosphoramidates are hydrolysed to d4T before exerting their antiviral activity.

## Introduction

The 2',3'-dideoxynucleoside (ddN) analogues, which are potent inhibitors of human immunodeficiency virus (HIV), act in their metabolic active 5'-triphosphate forms as substrates for reverse transcriptase (RT).<sup>1</sup> In essence, the ddN triphosphates (ddNTPs) compete with the natural substrates at the RT level for incorporation in DNA and pyrophosphate (PPi) functions as a leaving group.<sup>1,2</sup> In most cases, the bottleneck in the metabolic activation of dideoxynucleosides is the enzymatic phosphorylation of the modified nucleoside by cellular kinases. Therefore, a number of prodrug strategies have been developed to improve anti-HIV efficacy and reduce toxicity by delivering the modified nucleoside monophosphate into the cell.<sup>3–11</sup> It is worth noting that phosphoramidates of antiviral nucleosides are especially efficient as a nucleotide delivery system, particularly if an amino acid is used.<sup>4</sup> Among them, the phosphoramidate derivatives, such as a number of alkyl and aryl phosphoramidate mono-,<sup>5,6</sup> or di-esters<sup>7,8</sup> of AZT, d4T, and 3'-fluoro-dideoxythymidine (FLT), have shown promise as potential anti-HIV agents since they show enhanced antiviral activity and lower cytotoxicity when compared to the parent nucleoside. In these modifications the phosphate group is conjugated with hydrophobic amino acid esters, such as alanine, L-phenylalanine and L-tryptophan methyl ester. These pro-nucleotides are able to be internalized by PBMCs and CEM cells and further transformed to their monophosphates and corresponding triphosphates.<sup>10</sup> Additionally, ddN triphosphate analogues were also investigated for producing anti-ddNTP antibodies<sup>12</sup> and inhibiting RT-mediated drug resistance.<sup>13</sup>

In our continuous effort to search for amino acid derivatives as PPI mimics in enzymatic DNA synthesis,<sup>14–20</sup> 3-phosphono-L-alanine, L-aspartic acid, iminodiacetic acid and iminodipropionic acid proved to be efficient leaving groups when catalyzed by HIV-1 RT, suggesting efficient metal-chelation of these amino acid derivatives inside the polymerase active site. 3-Phosphono-L-Ala-dNMP (N = A, T and G) is efficiently incorporated in DNA resulting in 77–95% conversion to a P + 1 strand at 50  $\mu$ M after 60 min polymerase reaction by HIV-1 RT.<sup>19</sup> Therefore, we have studied the potential of the corresponding phosphoramidate dideoxynucleotide analogues to act as chain terminators. Hence, in the current study, d4T monophosphate was conjugated with 3-phosphono-L-alanine mono- or di-ester and the conjugates **1** and **2** (Fig. 1) were evaluated as chain terminators of HIV-1 RT. Additionally, L-Asp-d4TMP (**3**) and IDA-d4TMP (**4**) were synthesized and evaluated as substrates of HIV-1 RT. As well as ester derivatives (**1**, **2**), free acids (**3**, **4**) have been evaluated to investigate the structural requirements for cellular uptake of such derivatives. Among these analogues, 3-phosphono-L-alanine monoester phosphoramidate of d4TMP (**2**) proved to be efficient in single nucleotide incorporation and resulted in 75% conversion to P + 1 strand at 100  $\mu$ M after 60 min. Full chain termination can

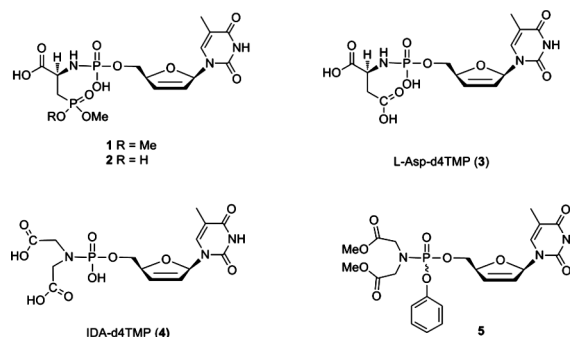


Fig. 1 Structure of phosphoramidate derivatives of d4T.

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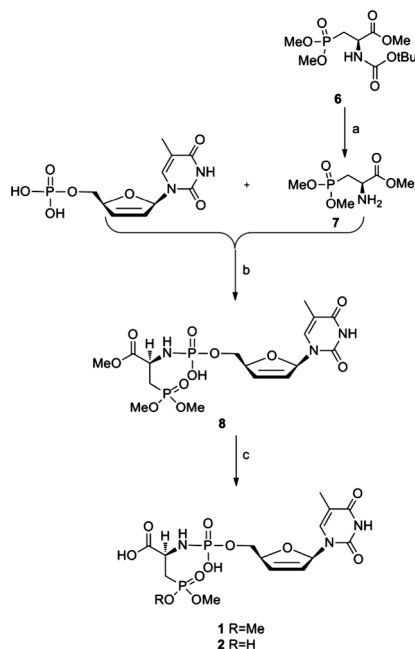
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be observed for compound **2** at higher substrate concentration (200  $\mu\text{M}$  to 1 mM). It was found that the steady-kinetics study of compound **2** demonstrated a typical Michaelis–Menten kinetic profile. In addition, to increase the hydrophobicity of the potential prodrugs, which is important for cellular uptake, the aryl phosphoramidate analogue **5** was synthesized. The selection of the phenol leaving group is based on the work of McGuigan and coworkers.<sup>4,7,8</sup> Compounds **1–5** were further tested for their anti-HIV activity in the cells.

## Results and discussion

### Synthesis of d4T phosphoramidate derivatives

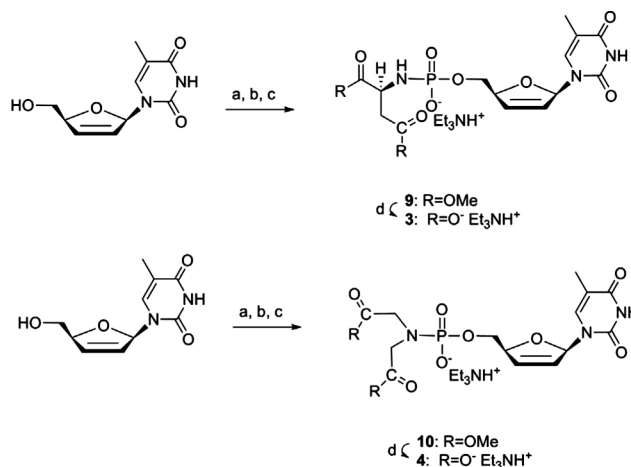
The synthesis of the amino acid phosphoramidate derivatives of d4T (**1–5**) was carried out by two synthetic routes. Compounds **1** and **2** were synthesized from d4T monophosphate (d4TMP) and 3-phosphono-L-alanine trimethyl ester (**7**) by an EDAC-mediated coupling, followed by deprotection in basic conditions. The synthesis of compounds **1** and **2** is shown in Scheme 1. The synthetic strategy starts by removal of the Boc protecting group of **6** with trifluoroacetic acid in  $\text{CH}_2\text{Cl}_2$ .<sup>21,22</sup> The resulting 3-phosphono-L-alanine trimethyl ester (**7**) can directly be used for further amidation. The monophosphate of d4T (d4TMP) was prepared by treatment of d4T with  $\text{POCl}_3$  as described before.<sup>23,24</sup> In order to avoid purification problems, this reaction was carried out using a small amount (120 mg) of d4T. The EDAC-mediated coupling of d4TMP with the primary amino function of **7** yielded the phosphoramidate methyl ester precursor (**8**) in 57.1%. Deprotection of both the carboxylic acid ester and phosphonate ester is accomplished in basic condition to give the desired products (**1** and **2**) in 78.6% overall yield. Column chromatographic purification followed by HPLC was used to purify the final products.



**Scheme 1** Reagents and conditions: (a)  $\text{CF}_3\text{COOH}$ ,  $\text{CH}_2\text{Cl}_2$ , rt, 4 h; (b) EDAC,  $\text{H}_2\text{O}$ , rt, 3 h; (c) 0.2 M NaOH in  $\text{MeOH}/\text{H}_2\text{O}$  1 : 2, rt, 4–24 h.

In addition, the hydrolysis of the phosphonate ester function of **8** was also investigated using trimethylsilyl bromide (TMSBr) as reagent in anhydrous  $\text{CH}_2\text{Cl}_2$ . The reaction failed even at 0  $^\circ\text{C}$ , and only degradation products were obtained (demonstrated by the isolation of the nucleobase thymine). This means that we were not able to obtain the fully deprotected congener of compound **8**.

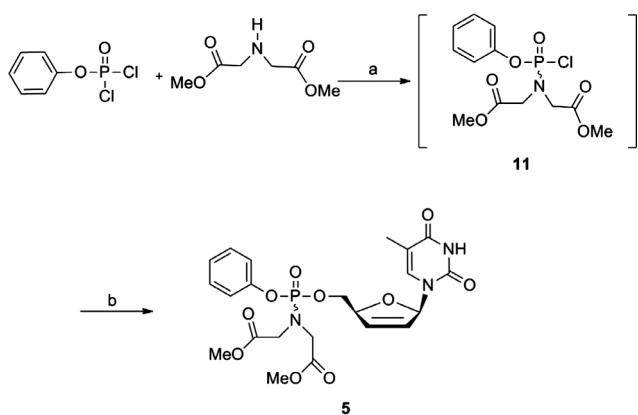
The second synthetic route used for the synthesis of the phosphoramidate derivatives of d4T is based on the one pot procedure described by Ludwig and Yoshikawa *et al.*<sup>25,26</sup> The synthesis of L-Asp and IDA phosphoramidate derivatives (**3** and **4**) of d4T is shown in Scheme 2. 2',3'-Didehydridideoxythymidine (d4T) was first phosphorylated and the resulting nucleoside phosphodichloridate was then reacted with the hydrochloride salt of the appropriate amino compound in the presence of tributylamine. After conversion to the triethylammonium salt, the resulted phosphoramidate compounds **9** and **10** were deprotected in basic media and subjected to ion exchange chromatography for purification, affording the final compounds **3** and **4** as triethylammonium salts in good yields (total 37.2–46.7%). As the phosphorylation of d4T requires anhydrous reaction conditions, this one-pot method is convenient when using the commercially available hydrochloride salt of amino acid ester.



**Scheme 2** Reagents and conditions: (a)  $\text{POCl}_3$ ,  $\text{PO}(\text{OMe})_3$ , 0  $^\circ\text{C}$ , 3 h; (b) (L-Asp) $\text{Me}_2\cdot\text{HCl}$  or (IDA) $\text{Me}_2\cdot\text{HCl}$ ,  $n\text{-Bu}_3\text{N}$ , DMF dry, 0  $^\circ\text{C}$  to rt, 3 min; (c) 1 M TEAB, rt, 30 min; (d) 0.4 M NaOH in  $\text{MeOH}/\text{H}_2\text{O}$  1 : 2, rt, 4 h.

### Synthesis of phenyl phosphoramidate derivative of d4T

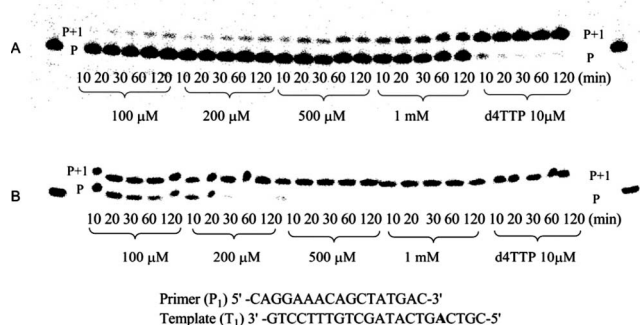
The synthesis of phenylphosphoramidate derivative (**5**) was achieved following Scheme 3, which is similar to the method described before for the synthesis of arylphosphoramidate derivatives of d4T.<sup>27,28</sup> In brief, phenylphosphorodichloride was condensed with iminodiacetate methylester hydrochloride to obtain the monochloridate (**11**). Treatment of monochloridate (**11**) in methylene chloride and triethylamine with d4T furnished the desired phenylphosphoramidate derivative **5**. Due to the stereochemistry of the phosphorus chiral center,<sup>9,11</sup> phenyl substituted phosphoramidate derivatives of d4T exist as a mixture of two possible diastereoisomers as evidenced from their  $^1\text{H}$ ,  $^{13}\text{C}$  and  $^{31}\text{P}$  NMR spectra. These isomers could not be separated by regular column chromatography and only fractions enriched with one of the other isomer were obtained.



**Scheme 3** Reagents and conditions: (a) TEA, dry  $\text{CH}_2\text{Cl}_2$ ,  $-78^\circ\text{C}$  to rt, 15 h; (b) d4T, rt, 5 d.

### Single primer extension by HIV-1 RT with d4T phosphoramidate derivatives (1–4) as substrates

As a key enzyme to replicate the single-stranded RNA genome and produce a double-stranded proviral DNA in HIV life-cycle, RT represents one of the main target enzymes in treating HIV infection. A high mutation rate with HIV-1 RT indicated its important flexibility and high tolerance towards chemically modified nucleotide substrates.<sup>29</sup> In previous studies,<sup>14–19</sup> it was shown that HIV-1 RT was able to accept amino acid dNMPs as substrate instead of natural dNTPs, during template-directed DNA synthesis. The potential to use this “new leaving group” concept for chain termination in DNA synthesis was evaluated using d4T as example and the amino acid derivatives 1–4. The ability of 3-phosphono-L-alanine di- or mono-methyl ester phosphoramidate analogues (1 and 2) to function as substrates for HIV-1 RT was investigated by the gel-based single nucleotide incorporation assay (Fig. 2). The natural dideoxynucleoside triphosphate (d4TTP) was used as reference.



**Fig. 2** Single incorporation of primer of  $P_1T_1$  (125 nM) by HIV-1 RT with phosphoramidate substrate concentrations and time intervals (min) as indicated,  $[\text{HIV-1 RT}] = 0.025 \text{ U } \mu\text{L}^{-1}$ . Panel A: incorporation of compound 1; panel B: incorporation of compound 2; d4TTP (10  $\mu\text{M}$ ) used as reference.

During a single nucleotide incorporation assay, compound 1 only resulted in 30% conversion to a P + 1 strand in 30 min at 1 mM. However, the use of compound 2 resulted in 75% incorporation in 60 min at 100  $\mu\text{M}$ . With higher concentrations (200  $\mu\text{M}$  to 1 mM), similar results were obtained and only P+1

product was formed. As expected, a single incorporated d4TMP functions as a chain terminator of HIV-1 RT. Compound 2 is 10-fold more efficient than compound 1 in the single primer-extension assay. This suggests that the 3-phosphono-L-alanine methyl ester can be better accommodated in the active site of the polymerase than the diester.

Under identical HIV-1 RT concentration, L-Asp-d4TMP (3) functioned as a suboptimal substrate: 20–25% P + 1 incorporation product was obtained after 120 min at 500  $\mu\text{M}$  concentration. High concentration of the substrate IDA-d4TMP (4) was required in order to incorporate d4TMP into DNA in a low yield (11–16% P + 1 incorporation product after 120 min at 1 mM concentration).

The efficiency of compound 2 was further investigated by determining the kinetic parameters  $K_m$  and  $K_{cat}$ .<sup>30,31</sup> Steady-state kinetic analysis (Table 1) indicates that the incorporation of compound 2 by HIV-1 RT results in 26-fold increase in the  $K_m$  value, and only 1.2-fold decrease in  $K_{cat}$  value, compared to those for d4TTP. Although the overall catalytic efficiency, the  $K_{cat}/K_m$  ratio, is decreased 31-fold, the  $K_{cat}$  value indicates the catalytic efficiency of HIV-1 RT for the phosphoramidate nucleotide (2) is only a little lower than for d4TTP. It should be mentioned that these kinetics measure the whole process of incorporation (including translocation and DNA release) and that the real incorporation kinetics may be underestimated.<sup>32</sup> Whatever, these data indicate efficient nucleophilic displacement of 3-phosphono-L-alanine monomethyl ester when the phosphoramidate is bound in the active site.

### Antiviral evaluation of phosphoramidate analogues (1–5) of dideoxynucleotides against HIV-1 and HIV-2

The antiviral activity of compounds 1–5 against HIV-1 and HIV-2 were evaluated in MT-4 cells, CEM/0 and CEM/TK<sup>-</sup> cells (Table 2). In all of these experiments, d4T and d4TMP were used as references. Compound 1 is 14 and 11-fold less active than compounds 2–4 in MT-4 cells and CEM/0 cells, respectively. Compound 4 is more efficient than compound 3 in CEM/0 cells. Compound 2 shows antiviral activity against HIV-1 and HIV-2 at concentrations of 0.47  $\mu\text{M}$  and 0.19  $\mu\text{M}$ , respectively, in MT-4 cells. Additionally, compound 2 has similar activity as d4T in CEM/0 cells. However, all of these d4T derivatives are much less active in CEM/TK<sup>-</sup>. These results indicate that the d4T phosphoramidate analogues and d4TMP itself were probably hydrolyzed to d4T in culture medium before penetration into the CEM/TK<sup>-</sup> cells.

In general, compounds 1–4 apparently serve as a source of d4T in cells and none of these phosphoramidate analogues possess marked cytotoxicity. The lower activity of compound 1 may be due to its less efficient hydrolysis to d4T than compound 2. In addition, compound 5 has a limited activity against HIV in MT-4 cells. Three mixtures consisting of varying ratios of the  $R_p$  and  $S_p$  isomers of the phosphotriester were evaluated for their anti-HIV activity. As we did not assign the absolute phosphorus stereochemistry, the isomer showing  $\delta 4.35$  ppm ( $^{31}\text{P}$  NMR) was called the upfield isomer and the other was the downfield isomer. The isomer mixtures denominated **5b** (ratio upfield/downfield = 2 : 1) and **5c** (ratio upfield/downfield = 1 : 1) were almost equipotent, whereas the mixture of isomers **5a** (ratio upfield/downfield = 1 : 2) was 12- and 4-fold less active than **5b** against HIV-1 and HIV-2,

**Table 1** Steady-state kinetics of single nucleotide incorporation into P<sub>1</sub>T<sub>1</sub> by HIV-1 RT

Compound	$V_{\max}$ (nM min <sup>-1</sup> )	$K_{\text{cat}}$ (min <sup>-1</sup> )	$K_m$ (μM)	$K_{\text{cat}}/K_m$ (×10 <sup>-3</sup> min <sup>-1</sup> μM <sup>-1</sup> )
d4TTP	7.92 ± 0.39 [HIV-1 RT] = 0.0125 U μL <sup>-1</sup> = 18.1 nM	0.44 ± 0.02	3.57 ± 0.53	123.2
Compound <b>2</b>	13.22 ± 1.08 [HIV-1 RT] = 0.025 U μL <sup>-1</sup> = 36.2 nM	0.37 ± 0.03	93.33 ± 16.55	3.96

**Table 2** Antiviral activity test of compounds **1–5** against HIV-1 and HIV-2

Compound	MT-4 cells <sup>a</sup>			CEM/0 <sup>b</sup>	CEM/TK <sup>-c</sup>	CEM/0 <sup>b</sup>
	IC <sub>50</sub> (μM) HIV-1	IC <sub>50</sub> (μM) HIV-2	CC <sub>50</sub> (μM)	IC <sub>50</sub> (μM) HIV-2	IC <sub>50</sub> (μM) HIV-2	CC <sub>50</sub> (μM)
<b>1</b>	4.26 ± 0.58	1.18 ± 0.29	>259	16.52 ± 1.11	258.64 ± 0.00	259
<b>2</b>	0.47 ± 0.26	0.19 ± 0.03	145.76 ± 4.73	1.44 ± 0.11	176.52 ± 28.65	266
<b>3</b>	0.36 ± 0.12	0.24 ± 0.05	169.70 ± 2.86	6.55 ± 4.88	183.66 ± 28.92	298
<b>4</b>	0.31 ± 0.02	0.21 ± 0.09	172.68 ± 3.72	2.13 ± 0.46	155.14 ± 19.99	298
<b>5a</b>	183.66 ± 28.92	81.39 ± 13.51	>239	>239	>239	>239
<b>5b</b>	14.90 ± 1.01	22.54 ± 0.80	>239	>239	>239	>239
<b>5c</b>	17.11 ± 0.63	19.04 ± 1.43	>239	>239	>239	>239
d4T	0.30 ± 0.03	0.19 ± 0.07	145.76 ± 4.73	3.02 ± 1.52	217.42 ± 11.07	268
d4TMP	0.39 ± 0.10	0.33 ± 0.10	187.05 ± 2.33	1.64 ± 0.48	170.11 ± 21.34	411

<sup>a</sup> HIV-1 (IIIB), HIV-2 (ROD). <sup>b</sup> CEM/0 cells (ROD). <sup>c</sup> CEM thymidine kinase deficient cells (ROD).

respectively. The diastereomeric excess was determined by <sup>31</sup>P NMR after high pressure liquid chromatography (HPLC). The observed differences in anti-HIV activity result from differences in hydrolytic stability of the different isomers. Most probably the poor anti-HIV activity of compound **5a–c** is due to the fact that both carboxylic acids are present as their methyl esters.

## Conclusion

In summary, the investigation of the synthesis and evaluation of the phosphoramidate analogues of d4TMP as chain terminators for HIV-1 RT has shown that compound **2** is a good substrate of HIV-1 RT resulting in 75% incorporation in 60 min at 100 μM. Compounds **1**, **3** and **4** are suboptimal substrates of HIV-1 RT and higher concentrations are needed to reach 11%–38% incorporation after 120 min. The capability to function as leaving groups in the enzymatic synthesis of DNA catalyzed by HIV-1 RT increase in the order of iminodiacetic acid < L-aspartic acid < 3-phosphono-L-alanine mono methylester, when conjugated with d4TMP.

In addition, compound **3** is 3-fold less efficient than compound **4** in CEM/0 cells. Compounds **2** and **4** exhibit similar anti-HIV activity as d4T and d4TMP in MT-4 cells and is 1.5–2-fold more efficient than d4T in CEM/0 cells. However, these compounds are less potent in CEM/TK<sup>-</sup> cells, as also observed for d4T and d4TMP. This suggests that d4TMP and the phosphoramidate analogues (**1–4**) are hydrolyzed to d4T in the extracellular medium or in an intracellular compartment. Indeed, following the McGuigan approach, the phosphoramidate diesters are taken up by cells and release the nucleoside monophosphate into the cell.<sup>7</sup> The negatively charged phosphoramidate monoesters, as produced by Wagner and coworkers,<sup>5</sup> can also be taken up by cells. The outcome of the phosphoramidate prodrug approach seems to be cell type and compound dependent. This, together with the different stability of the prodrugs in the extracellular medium, makes this approach difficult to predict.<sup>33</sup> In addition, compound

**5** lacks anti-HIV activity. This fully protected phosphoramidate seems to be resistant to enzymatic/chemical degradation to d4T in the given reaction circumstances. Preliminary results suggest a difference in hydrolytic stability of the different diastereomers of the phosphotriester in cell culture. The synthesis of new ester derivatives of the compounds mentioned in this article will be necessary to find a good balance between enzymatic stability, cellular uptake and intracellular processing.

## Experimental section

### General for chemical synthesis

For all reactions, analytical grade solvents were used. All moisture sensitive reactions were carried out in oven-dried glassware (135 °C). A Bruker Avance II 600 MHz or 500 MHz spectrometer and a Bruker Avance 300 MHz apparatus were used for <sup>1</sup>H NMR, <sup>13</sup>C NMR and <sup>31</sup>P NMR. All chemical shifts are listed in ppm. For sake of clarity of the NMR signal assignment, sugar protons and carbons are numbered with a prime. <sup>31</sup>P NMR chemical shifts are referenced to an external 85% H<sub>3</sub>PO<sub>4</sub> standard (δ = 0.000 ppm). UV spectra were recorded on a Varian Cary-300-Bio UV/Vis Spectrophotometer. Mass spectrometry was performed on a quadrupole orthogonal acceleration time-of-flight mass spectrometer (Q-ToF-2, Micromass, Manchester, UK) equipped with a standard electrospray probe (Z-spray, Micromass, Manchester, UK). Samples were dissolved in acetonitrile/water (1 : 1 v/v) and infused with a flow rate of 5 μL min<sup>-1</sup>. Electrospray capillary voltage was set to 3000 V and cone voltage to 30 V. Nitrogen gas was used for nebulization and desolvation. Accurate masses were determined by coinfusion of the samples with leucine enkephalin (YGGFL) and recalibration of the spectrum using the peak at *m/z* 556.2771 as lock mass. Precoated aluminum sheets (MN ALUGRAM SIL G/UV<sub>254</sub> 20 × 20 cm) were used for TLC; the spots were examined with UV light. Column chromatography was performed on ICN silica gel 63-200, 60 Å.



**Experimental procedure for the synthesis of intermediate 8 and compounds 1, 2: synthesis of 3-phosphono-L-alanine trimethyl ester (7).** (*S*)-methyl *N*-Boc-2-amino-3-(dimethoxyphosphoryl)propanoate (**6**) (0.84 g, 2.70 mmol) was synthesized according to the procedure described before by E. C. R. Smith *et al.*<sup>34</sup> It was dissolved in CH<sub>2</sub>Cl<sub>2</sub> (12 mL) and trifluoroacetic acid (3 mL, 40.39 mmol) was added in two portions in 1 h. The reaction mixture was stirred at room temperature for 4 h. The solvents were evaporated *in vacuo* and the obtained TFA salt was dissolved in MeOH (8 mL) and cooled to 0 °C. Then, the pH of solution was adjusted to 8 by adding triethylamine (2.2 mL). The solvents were removed *in vacuo* and the residue was purified by silica column chromatography (n-hexane/EtOAc/MeOH, 6 : 5 : 1 to 4 : 4 : 1). Then, the *N*-deprotected product **7** (0.43 g, 75.5%) was obtained as yellowish syrup and it can directly be used for the subsequent amidation reaction. TLC (CH<sub>2</sub>Cl<sub>2</sub>/CH<sub>3</sub>OH 9 : 1, v/v): *R*<sub>f</sub> 0.36; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 12.28 (br, 2H), 4.12 (m, 1H), 3.74–3.79 (m, 9H), 2.35–2.62 (m, 2H); <sup>31</sup>P NMR (121 MHz, CDCl<sub>3</sub>): δ 29.56; MS for C<sub>6</sub>H<sub>14</sub>N<sub>1</sub>O<sub>5</sub>P<sub>1</sub> (M + H)<sup>+</sup> calcd: 212.1 found: 211.9.

### Synthesis of d4TMP

To a solution of 3'-deoxy-2',3'-didehydrothymidine (100 mg, 0.45 mmol) in trimethylphosphate (1.25 mL) was added phosphoryl chloride (0.11 mL, 1.18 mmol) at 0 °C. The reaction mixture was stirred at room temperature for 1.5 h and another 0.11 mL of phosphoryl chloride was added. The reaction was continued to stir for 15 h and quenched by adding 1 M TEAB (pH 7.4) 15 mL. After the solvents were evaporated *in vacuo*, the residue was purified by silica column chromatography eluting with CHCl<sub>3</sub>, CHCl<sub>3</sub>/MeOH 5 : 1 to *i*-PrOH/NH<sub>3</sub>/H<sub>2</sub>O 7 : 1 : 1 to yield the desired d4TMP ammonium salt (55 mg, 36.5%) as white solid. <sup>1</sup>H NMR (300 MHz, D<sub>2</sub>O): δ 7.63 (s, 1H, H-6), 6.96 (m, 1H, H-1'), 6.51 (dd, *J*<sub>1</sub> = 1.6 Hz, *J*<sub>2</sub> = 6.0 Hz, 1H, H-2'), 5.91 (d, *J*<sub>1</sub> = 6.2, *J*<sub>2</sub> = 1.5 Hz, 1H, H-3'), 5.07 (m, 1H, H-4') 3.92 (m, 2H, H-5' and H-5''), 1.90 (q, 3H, -CH<sub>3</sub>); <sup>13</sup>C NMR (75 MHz, D<sub>2</sub>O): δ 166.34 (C-4), 151.89 (C-2), 137.76 (C-6), 134.16 (C-3'), 124.60 (C-2'), 111.10 (C-5), 89.70 (C-1'), 85.99 (d, <sup>3</sup>J(C, P) = 8.3 Hz, C-4'), 65.06 (d, <sup>2</sup>J(C, P) = 4.6 Hz, C-5'), 11.08 (CH<sub>3</sub>); <sup>31</sup>P NMR (121 MHz, D<sub>2</sub>O): δ 1.57 (ammonium salt) or 3.36 (triethylamine salt); MS for C<sub>10</sub>H<sub>13</sub>N<sub>2</sub>O<sub>7</sub>P<sub>1</sub> (M-H)<sup>-</sup> calcd: 303.0, found: 302.6.

### Coupling reaction

2',3'-Dideoxy-2',3'-didehydrothymidine-5'-monophosphate ammonium salt (50 mg, 0.15 mmol) and the crude 3-phosphono-L-alanine trimethyl ester (103 mg, 0.49 mmol) were suspended in 1 mL water under Argon. Then, *N*-ethyl-*N'*-(3-dimethylamino-propyl)-carbodiimide hydrochloride (EDAC, 94 mg, 0.49 mmol) was added to the suspension, and the reaction was stirred for 2 h at room temperature. The progress of the reaction was monitored by TLC (CHCl<sub>3</sub>/MeOH/H<sub>2</sub>O 5 : 3 : 0.5). The reaction mixture was evaporated in vacuum at 30 °C and further lyophilized to dryness. The residue was purified by silica column chromatography eluting with CHCl<sub>3</sub>, CHCl<sub>3</sub>/MeOH = 5 : 1, CHCl<sub>3</sub>/MeOH/H<sub>2</sub>O = 5 : 2 : 0.25 to yield compound **8** (42 mg, 57.1%) as yellowish solid. TLC (*i*-PrOH/NH<sub>3</sub>/H<sub>2</sub>O 7 : 1 : 2, v/v): *R*<sub>f</sub> 0.62; <sup>1</sup>H NMR (300 MHz, D<sub>2</sub>O): δ 7.64 (s, 1H, H-6), 6.99 (m, 1H, H-1'), 6.47 (dd, *J*<sub>1</sub> =

6.1, *J*<sub>2</sub> = 1.8 Hz, 1H, H-2'), 5.92 (dd, *J*<sub>1</sub> = 6.7, *J*<sub>2</sub> = 1.8 Hz, 1H, H-3'), 5.08 (m, 1H, H-4'), 3.94 (m, 2H, H-5' and H-5''), 3.68–3.80 (m, 9H, 3 OMe), 3.54 (m, 1H, -CHCOO), 2.28–2.36 (m, 2H, -CH<sub>2</sub>PO<sub>3</sub>), 1.90 (q, 3H, -CH<sub>3</sub>); <sup>13</sup>C NMR (75 MHz, D<sub>2</sub>O): δ 174.15 (d, <sup>3</sup>J(C, P) = 5.1 Hz, -COOH), 166.33 (C-4), 151.88 (C-2), 137.96 (C-6), 134.19 (C-2'), 124.80 (C-2'), 110.97 (C-5), 89.61 (d, <sup>3</sup>J(C, P) = 12.8 Hz, C-4'), 85.68 (C-1'), 64.68 (d, <sup>2</sup>J(C, P) = 5.1 Hz, C-5'), 52.59 (d, <sup>2</sup>J(C, P) = 3.0 Hz, -CHCOOH), 52.49 (d, <sup>2</sup>J(C, P) = 5.7 Hz, POCH<sub>3</sub>), 49.60 (COOCH<sub>3</sub>), 28.27 (d, <sup>1</sup>J(C, P) = 140.1 Hz, -CH<sub>2</sub>PO<sub>3</sub>), 11.10 (CH<sub>3</sub>); <sup>31</sup>P NMR (121 MHz, D<sub>2</sub>O): δ 31.86 (-CH<sub>2</sub>PO<sub>3</sub>), 4.79 (*N*-PO<sub>3</sub>); MS for C<sub>16</sub>H<sub>25</sub>N<sub>3</sub>O<sub>11</sub>P<sub>2</sub> (M-H)<sup>-</sup> calcd: 496.1, found: 495.8.

### Deprotection reaction

To the obtained ester intermediate **8** (39 mg, 0.08 mmol) was added 6 mL 0.2 M NaOH in MeOH/H<sub>2</sub>O 1 : 2 and the mixture were stirred at room temperature. The progress of the reaction was monitored by TLC (*i*-PrOH/NH<sub>3</sub>/H<sub>2</sub>O 7 : 1 : 2) until the disappearance of ester. The hydrolysis is stopped after 4 h and 24 h, respectively, to give the desired products as **1** and **2**. The reaction mixture was neutralized by addition of 1 M triethylammonium acetate (TEAA). The solvent was evaporated to dryness in vacuum. The residue was purified by silica column chromatography eluting with CHCl<sub>3</sub>, CHCl<sub>3</sub>/MeOH = 5 : 1, CHCl<sub>3</sub>/MeOH/H<sub>2</sub>O = 5 : 2.5 : 0.4, yielding compound **1** (18 mg, 46.6%), then *i*-PrOH/NH<sub>3</sub>/H<sub>2</sub>O (7 : 1 : 1 to 7 : 1 : 1.5) and compound **2** (12 mg, 32.0%) as white solid. An analytical sample was obtained by purification with preparative HPLC (waters 1525-2487 system) with a gradient of CH<sub>3</sub>CN in 50 mM TEAB buffer (pH = 7.4) on using Prep C18 5 μm column 19 × 150 mm at the flow rate of 3 mL min<sup>-1</sup>.

**2',3'-Dideoxy-2',3'-didehydrothymidine-5'-[3-(dimethoxyphosphoryl)-L-alanine] phosphoramidate (1).** TLC (*i*-PrOH/NH<sub>3</sub>/H<sub>2</sub>O 7 : 1 : 2, v/v): *R*<sub>f</sub> 0.53; UV: (H<sub>2</sub>O) λ<sub>max</sub> = 266 nm (log ε = 3.97); <sup>1</sup>H NMR (300 MHz, D<sub>2</sub>O, 5 °C): δ 7.70 (s, 1H, H-6), 6.97 (m, 1H, H-1'), 6.48 (ddd, *J*<sub>1</sub> = 6.1 Hz, *J*<sub>2</sub> = 1.8 Hz, *J*<sub>3</sub> = 1.8 Hz, 1H, H-2'), 5.92 (dd, *J*<sub>1</sub> = 6.0 Hz, *J*<sub>2</sub> = 1.8 Hz, *J*<sub>3</sub> = 1.8 Hz, 1H, H-3'), 5.07 (m, 1H, H-4'), 3.97 (m, 2H, H-5' and H-5''), 3.70–3.75 (m, 6H, 2 OMe), 3.57 (m, 1H, -CHCOO), 2.23–2.35 (m, 2H, -CH<sub>2</sub>PO<sub>3</sub>), 1.92 (q, 3H, -CH<sub>3</sub>); <sup>13</sup>C NMR (75 MHz, D<sub>2</sub>O, 5 °C): δ 178.14 (d, <sup>3</sup>J(C, P) = 9.0 Hz, -COOH), 166.42 (C-4), 151.89 (C-2), 137.92 (C-6), 134.10 (C-3'), 124.86 (C-2'), 111.24 (C-5), 89.61 (C-1'), 85.85 (C-4'), 65.10 (d, <sup>2</sup>J(C, P) = 5.3 Hz, C-5'), 52.64 (dd, <sup>2</sup>J<sub>1</sub>(C, P) = 2.0 Hz, <sup>2</sup>J<sub>2</sub>(C, P) = 6.8 Hz, -CHCOOH), 51.87 (d, <sup>2</sup>J(C, P) = 3.4 Hz, POCH<sub>3</sub>), 29.15 (d, <sup>1</sup>J(C, P) = 135.6 Hz, -CH<sub>2</sub>PO<sub>3</sub>), 11.10 (CH<sub>3</sub>); <sup>31</sup>P NMR (121 MHz, D<sub>2</sub>O): δ 33.90 (-CH<sub>2</sub>PO<sub>3</sub>), 5.66 (*N*-PO<sub>3</sub>); HRMS for C<sub>15</sub>H<sub>23</sub>N<sub>3</sub>O<sub>11</sub>P<sub>2</sub> (M-H)<sup>-</sup> calcd: 482.0735, found: 482.0738.

**2',3'-Dideoxy-2',3'-didehydrothymidine-5'-[3-(methoxyphosphoryl)-L-alanine] phosphoramidate (2).** TLC (*i*-PrOH/NH<sub>3</sub>/H<sub>2</sub>O 7 : 1 : 2, v/v): *R*<sub>f</sub> 0.40; UV: (H<sub>2</sub>O) λ<sub>max</sub> = 266 nm (log ε = 3.97); <sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O, 5 °C): δ 7.62 (s, 1H, H-6), 6.90 (m, 1H, H-1'), 6.45 (d, *J* = 6.0 Hz, 1H, H-2'), 5.89 (m, 1H, H-3'), 5.12 (m, 1H, H-4'), 3.92 (m, 2H, H-5' and H-5''), 3.61 (m, 1H, -CHCOOH), 3.45 (q, 3H, OMe), 1.86–1.99 (m, 2H, -CH<sub>2</sub>PO<sub>3</sub>), 1.84 (q, 3H, -CH<sub>3</sub>); <sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O, 5 °C): δ 181.9 (-COOH), 169.63 (C-4), 155.02 (C-2), 141.05 (C-6), 137.28 (C-3'), 127.53 (C-2'), 114.25 (C-5), 92.54 (C-1'), 88.92 (d, <sup>3</sup>J(C, P) = 9.1 Hz,

C-4'), 67.60 (d,  $^2J(C, P) = 4.8$  Hz, C-5'), 55.73 (d,  $^2J(C, P) = 18.1$  Hz, -CHCOOH), 53.98 (d,  $^2J(C, P) = 4.8$  Hz, POCH<sub>3</sub>), 33.87 (d,  $^1J(C, P) = 130.5$  Hz,  $^3J(C, P) = 7.5$  Hz, -CH<sub>2</sub>PO<sub>3</sub>), 14.21 (CH<sub>3</sub>);  $^{31}P$  NMR (202 MHz, D<sub>2</sub>O):  $\delta$  22.0 (-CH<sub>2</sub>PO<sub>3</sub>), 6.10 (N-PO<sub>3</sub>); HRMS for C<sub>14</sub>H<sub>21</sub>N<sub>3</sub>O<sub>11</sub>P<sub>2</sub> (M-H)<sup>-</sup> calcd: 468.0578, found: 468.0600.

**General procedure for synthesis of intermediates 9, 10 and compounds 3, 4: 2',3'-dideoxy-2',3'-dideohydrothymidine-5'-(dimethyl-L-aspartate)phosphoramidate triethylammonium salt (9, intermediate for compound 3)**

All solids were dried in a desiccator under high vacuum one day prior to reaction. The nucleoside (50 mg, 0.22 mmol) was placed under argon atmosphere and dissolved in trimethylphosphate (500  $\mu$ L, 50  $\mu$ L/10 mg nucleoside). Upon cooling down to 0 °C, phosphorous trichloride (42  $\mu$ L, 0.44 mmol, 2 eq.) was added slowly *via* syringe. The resulting solution was stirred for 3 h at 0 °C and the reaction was monitored by TLC (*i*-PrOH/NH<sub>3</sub>/H<sub>2</sub>O 6:3:1, v/v). Upon total conversion of the starting nucleoside, a solution of dimethyl L-aspartic acid hydrochloride (434 mg, 2.2 mmol, 10 eq.) and n-tributylamine (500  $\mu$ L, 30 eq.) in dried DMF (2.2 mL) was added *via* syringe. The resulting mixture was stirred vigorously for 3 min and pipetted into a cold TEAB 1 M solution (5 mL). The resulting clear mixture was further stirred at room temperature for 30 min. Upon completion, all volatiles were evaporated under high vacuum. The product was purified by flash chromatography using a gradient of methanol and water in chloroform (5:1:0, 5:2:0.25, 5:3:0.5). The corresponding fractions were evaporated, lyophilized and the final compound (70 mg, colourless solid, yield 73%) was stored at -20 °C.

TLC (*i*-PrOH/NH<sub>3</sub>/H<sub>2</sub>O 6:3:1, v/v) *R*<sub>f</sub> 0.77;  $^1H$  NMR (D<sub>2</sub>O, 300 MHz):  $\delta$  7.56 (s, 1H, H-6), 6.89 (s, 1H, H-1'), 6.41 (s, 1H, *J* = 5.97 Hz, H-2'), 5.86 (s, 1H, *J* = 5.37 Hz, H-3'), 5.00 (bs, 1H, H-4'), 3.97 (m, 1H, H<sub>a</sub>), 3.88 (bs, 2H, H-5'), 3.70, 3.65 (2 s, 2  $\times$  3H, CH<sub>3</sub>-O), 3.13 (q, 6H, N-CH<sub>2</sub>-CH<sub>3</sub>, Et<sub>3</sub>N<sup>+</sup>), 2.72–2.70 (m, 2H, H <sub>$\beta$</sub> ), 1.83 (s, 3H, CH<sub>3</sub>), 1.21 (t, 9H, N-CH<sub>2</sub>-CH<sub>3</sub>, Et<sub>3</sub>N<sup>+</sup>) ppm.  $^{13}C$  NMR (D<sub>2</sub>O, 75 MHz):  $\delta$  173.04 (CO Asp 2x), 166.57 (C-4T), 152.16 (C-2T), 138.19 (C-6), 134.05 (C-2'), 125.24 (C-3'), 111.25 (C-5'), 89.75 (C-1'), 85.58 (C-4', *J* = 8.71 Hz), 64.71 (C-5'), 52.80 (CH<sub>3</sub>-O), 50.93 (CH<sub>3</sub>-O), 43.21 (Et<sub>3</sub>N<sup>+</sup>), 38.20 (CH<sub>a</sub>), 35.48 (CH <sub>$\beta$</sub> ), 11.27 (CH<sub>3</sub>-T), 8.14 (Et<sub>3</sub>N<sup>+</sup>) ppm.  $^{31}P$  NMR (D<sub>2</sub>O, 121 MHz):  $\delta$  5.59 ppm. HRMS (negative ionization): calculated for C<sub>16</sub>H<sub>21</sub>N<sub>3</sub>O<sub>10</sub>P<sup>-</sup>: 446.0970, found: 446.0966.

**2',3'-Dideoxy-2',3'-dideohydrothymidine-5'-(L-aspartyl)phosphoramidate triethylammonium salt (3).** The previously prepared compound (70 mg) was dissolved in 0.4 M sodium hydroxide solution in water : methanol 2:1 (1 mL) and stirred at room temperature until complete deprotection was achieved. Solvents were evaporated and the residue was submitted to preparative ion exchange chromatography (Source 15Q, Pharmacia) running a gradient of TEAB in water (0 to 30% in 15 min). The final compound was obtained as a white solid (yield 51–64%) and stored at -20 °C.

$^1H$  NMR (D<sub>2</sub>O, 300 MHz):  $\delta$  7.58 (s, 1H, H-6), 6.88 (s, 1H, H-1'), 6.39 (s, 1H, *J* = 6.09 Hz, H-2'), 5.84 (s, 1H, *J* = 6.06 Hz, H-3'), 5.00 (bs, 1H, H-4'), 3.90 (bs, 2H, H-5'), 3.70–3.67 (m, 1H, H<sub>a</sub>), 3.14 (q, 6H, N-CH<sub>2</sub>-CH<sub>3</sub>, Et<sub>3</sub>N<sup>+</sup>), 2.61–2.58 (m, 2H, H <sub>$\beta$</sub> ), 1.83 (s, 3H, CH<sub>3</sub>), 1.21 (t, 9H, N-CH<sub>2</sub>-CH<sub>3</sub>, Et<sub>3</sub>N<sup>+</sup>) ppm.  $^{13}C$  NMR (D<sub>2</sub>O, 125 MHz):  $\delta$  178.24 (CO Asp), 176.08 (CO Asp), 166.32 (C-4T),

151.85 (C-2T), 137.87 (C-6), 134.05 (C-2'), 124.81 (C-3'), 111.10 (C-5), 89.50 (C-1'), 85.70 (C-4', *J* = 9.65 Hz), 64.52 (C-5'), 46.30 (Et<sub>3</sub>N<sup>+</sup>), 39.70 (CH<sub>a</sub>), 34.75 (CH <sub>$\beta$</sub> ), 11.13, 7.86 (Et<sub>3</sub>N<sup>+</sup>) ppm.  $^{31}P$  NMR (D<sub>2</sub>O, 121 MHz):  $\delta$  6.34 ppm. HRMS (negative ionization): calculated for C<sub>14</sub>H<sub>17</sub>N<sub>3</sub>O<sub>10</sub>P<sup>-</sup>: 418.0657, found: 418.0659.

**2',3'-Dideoxy-2',3'-dideohydrothymidine-5'-(dimethyliminodiacetate)phosphoramidate triethylammonium salt (10, intermediate for compound 4).** The protocol for the synthesis of **10** is identical to the previously described method for the synthesis of the phosphoramidate (**9**) using a solution of dimethyl iminodiacetic acid hydrochloride (434 mg, 2.2 mmol, 10 eq.) and n-tributylamine (500  $\mu$ L, 30 eq.) in dried DMF (2.2 mL) in the second step. The corresponding compound **10** (74 mg, colourless solid, yield 76%) was obtained and stored at -20 °C.

TLC (*i*-PrOH/NH<sub>3</sub>/H<sub>2</sub>O 6:3:1, v/v) *R*<sub>f</sub> 0.50;  $^1H$  NMR (D<sub>2</sub>O, 300 MHz):  $\delta$  7.52 (s, 1H, H-6), 6.85 (s, 1H, H-1'), 6.39 (s, 1H, *J* = 5.97 Hz, H-2'), 5.83 (s, 1H, *J* = 5.58 Hz, H-3'), 4.95 (bs, 1H, H-4'), 3.80–3.60 (m, 5H, H-5' and 2  $\times$  CH<sub>2</sub>), 3.57 (bs, 6H, CH<sub>3</sub>-O), 3.11 (q, 6H, N-CH<sub>2</sub>-CH<sub>3</sub>, Et<sub>3</sub>N<sup>+</sup>), 1.82 (s, 3H, CH<sub>3</sub>) 1.19 (t, 9H, N-CH<sub>2</sub>-CH<sub>3</sub>, Et<sub>3</sub>N<sup>+</sup>) ppm.  $^{13}C$  NMR (D<sub>2</sub>O, 126 MHz):  $\delta$  173.83 (2  $\times$  CO Ac), 166.62 (C-4T), 152.144 (C-2T), 138.44 (C-6), 134.62 (C-2'), 124.73 (C-3'), 111.06 (C-5'), 89.70 (C-1'), 85.85 (C-4, *J* = 10.5 Hz), 64.85 (C-5'), 52.13 (CH<sub>2</sub> IDA), 48.69 (CH<sub>3</sub>-O), 46.58 (Et<sub>3</sub>N<sup>+</sup>), 11.37, 8.16 (Et<sub>3</sub>N<sup>+</sup>) ppm.  $^{31}P$  NMR (D<sub>2</sub>O, 121 MHz):  $\delta$  6.45 ppm. HRMS (negative ionization): calculated for C<sub>16</sub>H<sub>21</sub>N<sub>3</sub>O<sub>10</sub>P<sup>-</sup>: 446.0970, found: 446.0976.

**2',3'-Dideoxy-2',3'-dideohydrothymidine-5'-iminodiacetate phosphoramidate triethylammonium salt (4).** The deprotection of compound **10** (74 mg) is carried out in the same way as the deprotection of **9**. The final compound **4** was obtained as a white solid (yield 51–60%) and stored at -20 °C.

$^1H$  NMR (D<sub>2</sub>O, 300 MHz):  $\delta$  7.51 (s, 1H, H-6), 6.85 (s, 1H, H-1'), 6.38 (s, 1H, *J* = 6.18 Hz, H-2'), 5.84 (s, 1H, *J* = 5.58 Hz, H-3'), 4.97 (bs, 1H, H-4'), 3.91–3.88 (m, 5H, H-5'), 3.67, 3.64 (2 s, 4H, 2  $\times$  CH<sub>2</sub>), 3.11 (q, 6H, N-CH<sub>2</sub>-CH<sub>3</sub>, Et<sub>3</sub>N<sup>+</sup>), 1.81 (s, 3H, CH<sub>3</sub>), 1.19 (t, 9H, N-CH<sub>2</sub>-CH<sub>3</sub>, Et<sub>3</sub>N<sup>+</sup>) ppm.  $^{13}C$  NMR (D<sub>2</sub>O, 151 MHz):  $\delta$  178.91 (2  $\times$  CO Ac), 166.42 (C-4), 151.94 (C-2), 137.88 (C-6), 134.00 (C-2'), 124.81 (C-3'), 111.14 (C-5'), 89.62 (C-1'), 85.60 (C-4', *J* = 9.50 Hz), 64.92 (C-5'), 52.96 (CH<sub>2</sub> IDA), 46.37 (Et<sub>3</sub>N<sup>+</sup>), 11.16, 8.10 (Et<sub>3</sub>N<sup>+</sup>) ppm.  $^{31}P$  NMR (D<sub>2</sub>O, 121 MHz):  $\delta$  6.97 ppm. HRMS (negative ionization): calculated for C<sub>14</sub>H<sub>17</sub>N<sub>3</sub>O<sub>10</sub>P<sup>-</sup>: 418.0657, found: 418.0663.

**2',3'-Dideoxy-2',3'-dideohydrothymidine-5'-(phenyl dimethyliminodiacetate phosphate) (5).** In a dry flask, iminodiacetate methylester hydrochloride (0.99 g, 5 mmol) and dry CH<sub>2</sub>Cl<sub>2</sub> (37.5 mL) were added and cooled to -78 °C. Then phenyl dichlorophosphate (1.05 g, 0.74 mL, 5 mmol) was added into the flask. The reaction mixture were stirred and kept at -78 °C. Anhydrous triethylamine (2.1 mL, 15 mmol) was added dropwise at -78 °C. After 15 min, the reaction mixture was left to warm up to room temperature and stirred overnight. The formation of the phosphonochloridate was monitored by  $^{31}P$  NMR and an additional portion of triethylamine (2.1 mL, 15 mmol) was added to the reaction mixture and the contents were allowed to stir. After stirring at room temperature for 2 h, d4T (0.45 g, 2.0 mmol) was added to the above reaction flask and the reaction mixture was stirred at room temperature for 5 days. The suspension was filtered

and the filtrate was washed with anhydrous  $\text{CH}_2\text{Cl}_2$   $2 \times 25$  mL. Solvent was removed under reduced pressure. The crude reaction product was purified by column chromatography (100%  $\text{CHCl}_3$  to 5% MeOH in  $\text{CHCl}_3$ ) to yield compound **5** as yellowish syrup (0.9 g, 86.0%). Preparative HPLC (waters 1525–2487 system) with a gradient of  $\text{CH}_3\text{CN}$  in  $\text{H}_2\text{O}$  on C18 5  $\mu\text{m}$  column  $19 \times 150$  mm was used to purify the syrup. Sample **5a**, **5b** and **5c** were obtained as (not assigned) fractions of  $R_p$  and  $S_p$  with the ratio of 1 : 2, 2 : 1 and 1 : 1, determined by the relative intensity of two peaks of  $^{31}\text{P}$  NMR ( $\delta$  4.35, 5.11 ppm).

TLC ( $\text{CHCl}_3/\text{CH}_3\text{OH}$  18 : 1, v/v):  $R_f$  0.35–0.40;  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ,  $5^\circ\text{C}$ ):  $\delta$  8.20 (s, 1H, NH), 7.34–7.36 (m, 2H, phenyl), 7.32 (s, 1H, H-6), 7.17–7.21 (m, 3H, phenyl), 7.03 (m, 1H, H-1'), 6.34 (m, 1H, H-2'), 5.88 (m, 1H, H-3'), 5.04 (m, 1H, H-4'), 4.37 (m, 2H, H-5' and H-5''), 3.88–3.98 (m, 4H, 2- $\text{CH}_2$ ), 3.67 (s, 6H, 2- $\text{OCH}_3$ ), 1.78 (q, 3H,  $-\text{CH}_3$ );  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ ,  $5^\circ\text{C}$ ):  $\delta$  170.20 ( $-\text{COOCH}_3$ ), 163.41 (C-4), 150.52 (C-2), 135.67 (C-6), 133.19 (C-2'), 129.68 (C-phenyl), 126.93 (C-3'), 125.22 (C-phenyl), 120.15 (C-phenyl), 111.27 (C-5), 89.29 (C-1'), 84.48 (d,  $^3\text{J}(\text{C}, \text{P}) = 8.8$  Hz, C-4'), 65.98 (d,  $^2\text{J}(\text{C}, \text{P}) = 3.1$  Hz, C-5'), 52.24 ( $-\text{OCH}_3$ ), 47.44 ( $-\text{CH}_2$ ), 47.41 ( $-\text{CH}_2$ ), 12.35 ( $\text{CH}_3$ );  $^{31}\text{P}$  NMR (202 MHz,  $\text{CDCl}_3$ ):  $\delta$  4.35, 5.11; HRMS for  $\text{C}_{22}\text{H}_{26}\text{N}_5\text{O}_{10}\text{P}$  (M-H) $^-$  calcd: 522.1288, found: 522.1281.

### Enzymatic synthesis

**Oligodeoxyribonucleotides.** DNA oligonucleotides  $P_1$  and  $T_1$  were purchased from Sigma Genosys and Eurogentec. The concentrations were determined with a Varian Cary-300-Bio UV Spectrophotometer. The lyophilized oligonucleotides were dissolved in diethylpyrocarbonate (DEPC)-treated water and stored at  $-20^\circ\text{C}$ . The primer oligonucleotides were 5'-labeled with [ $\gamma$ - $^{33}\text{P}$ ] ATP (Perkin Elmer) using T4 polynucleotide kinase (New England Biolabs) according to the standard procedures. Labeled oligonucleotides were further purified with Illustra<sup>TM</sup> Microspin<sup>TM</sup> G-25 columns (GE Healthcare).

**DNA polymerase reactions.** 5'-End-labeled primer was annealed to its template by combining primer and template at a molar ratio of 1 : 2 and heating the mixture to  $70^\circ\text{C}$  for 10 min, followed by slow cooling to room temperature over a period of 2 h. The complex of  $P_1T_1$  was used for the single nucleotide incorporation study of compounds **1–4**. A series of 20- $\mu\text{L}$ -batch reactions were performed for the enzyme HIV-1 RT (Ambion, Inc.;  $10 \text{ U } \mu\text{L}^{-1}$  stock solutions). The final mixture with HIV-1 RT contained 125 nM primer-template complex, RT buffer (50 mM Tris-HCl, 50 mM KCl, 10 mM  $\text{MgCl}_2$ , 0.5 mM spermidine, 10 mM DTT; pH 8.3), 0.025 U/ $\mu\text{L}$  HIV-1 RT, and different concentrations of phosphoramidate substrates. Mixtures were incubated at  $37^\circ\text{C}$ , and aliquots (2.5  $\mu\text{L}$ ) were removed and quenched after 10, 20, 30, 60, 120 min. In the control reaction with the natural nucleotides,  $10 \mu\text{M}$  d4TTP was used.

**Polyacrylamide electrophoresis.** All polymerase reaction (2.5  $\mu\text{L}$ ) were quenched by the addition of 10  $\mu\text{L}$  of loading buffer (90% formamide, 0.05% bromophenol blue, 0.05% xylene cyanol, and 50 mM ethylenediaminetetraacetic acid (EDTA)). Samples were heated at  $75^\circ\text{C}$  for 5 min prior to analysis by electrophoresis for 2.5 h at 2000 V on a 0.4-mm 20% denaturing gel in the presence of a 100 mM Tris-borate, 2.5 mM EDTA buffer, pH

8.3. Products were visualized by phosphor imaging. The amount of radioactivity in the bands corresponding to the products of enzymatic reactions was determined with the imaging device Cyclone and the associated Optiquant image analysis software (Perkin-Elmer).

**Steady-state kinetics of single nucleotide incorporation.** To determine the kinetic parameters for the incorporation of compound **2** and a natural nucleoside d4TTP, a steady-state kinetics assay was carried out. The reaction was started by adding HIV-1 RT to  $P_1$ - $T_1$  complex, RT buffer, compound **2** and d4TTP. The final mixture (20  $\mu\text{L}$ ) contained 125 nM primer-template complex, and various concentrations of compound **2** or d4TTP, buffer, 0.025 U  $\mu\text{L}^{-1}$  HIV-1 RT for **5** and 0.0125 U  $\mu\text{L}^{-1}$  HIV-1 RT for d4TTP, respectively. The range of concentrations for phosphoramidates was optimized according to a  $K_m$  value for the incorporation of an individual nucleotide. Enzymatic reactions were carried out to attain 5–25% incorporation with appropriate substrate concentrations (5–200  $\mu\text{M}$  used for compound **2** and 1–20  $\mu\text{M}$  used for d4TTP) incubated at  $37^\circ\text{C}$  and run for 8–10 different time intervals (1–10 min). The amount of primer template (125 nM) was 3.5-fold excess of HIV-1 RT (36.2 nM). Products were separated on a 20% polyacrylamide gel and quantitated by phosphor imaging. The incorporation velocities were calculated based on the percentage of single-nucleotide extension product (P + 1 band). The kinetic parameters ( $V_{\text{max}}$  and  $K_m$ ) were determined by plotting  $V$  ( $\text{nM min}^{-1}$ ) versus substrate concentration ( $\mu\text{M}$ ) and fitting the data point to a nonlinear Michaelis–Menten regression using GraphPad Prism software. The  $K_{\text{cat}}$  was calculated according to the used enzyme concentration.

### Biological evaluation of phosphoramidate analogues of dideoxynucleotides against HIV-1 and HIV-2

The biological evaluation of phosphoramidate analogues of d4T against HIV-1 and HIV-2 was carried out including d4T and its 5'-*O*-monophosphate as references.

The methodology of the anti-HIV assays was as follows. The anti-HIV activity and cytotoxicity of the compounds were evaluated against wild-type HIV-1 strain III<sub>B</sub> and HIV-2 strain ROD in MT-4 cell cultures using the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) method.<sup>35</sup> Briefly, virus stocks were titrated in MT-4 cells and expressed as the 50% cell culture infective dose (CCID<sub>50</sub>). MT-4 cells were suspended in culture medium at  $1 \times 10^5$  cells  $\text{mL}^{-1}$  and infected with HIV at a multiplicity of infection of 0.02. Immediately after viral infection, 100  $\mu\text{L}$  of the cell suspension was placed in each well of a flat-bottomed microtiter tray containing various concentrations of the test compounds. The test compounds were dissolved in DMSO at 50 mM or higher. After 4 days of incubation at  $37^\circ\text{C}$ , the number of viable cells was determined using the MTT method. Compounds were tested in parallel for cytotoxic effects in uninfected MT-4 cells.

CEM cells were infected as previously described. Briefly,  $4 \times 10^5$  cells  $\text{mL}^{-1}$  were infected with HIV-2 at  $\approx 100$  CCID<sub>50</sub> (50% cell culture infective dose) per mL of cell suspension. The thymidine kinase-deficient CEM (CEM/TK<sup>-</sup>) cell cultures were also infected with HIV-2. Then, 100  $\mu\text{L}$  of the infected cell suspensions was transferred into 96-well microtiter plate wells and mixed with 100  $\mu\text{L}$  of the appropriate dilutions of the test compounds. After 4 days



of incubation at 37 °C, the giant cell formation was recorded microscopically in the HIV-infected cell cultures.

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